



INSTITUTO DE HIGIENE E
MEDICINA TROPICAL
DESDE 1902

GENETICS OF DRUG RESISTANCE IN PARASITOLOGY

CU characterization:

CU name:

Genetics of drug resistance in parasitology

Scientific area acronym:

PA

Duration:

Semestral

Working hours:

56

Contact hours:

31

ECTS:

3

Observations:

Opcional CU

Teacher in charge and respective teaching load in the CU:

Fátima Nogueira - 9 hours

Other teachers and respective teaching load in the CU:

Sandra Antunes – 5.4 hours

Ana Domingos – 5.4 hours

Carla Sousa – 4.3 hours

João Pinto – 3.2 hours

Sofia Cortes – 3.2 hours

Olga Matos – 4.3 hours

Isabel Maurício – 5.4 hours

Manuela Calado – 5.4 hours

Docentes convidados – 9 hours

GENETICS OF DRUG RESISTANCE IN PARASITOLOGY

Intended learning outcomes (knowledge, skills and competences to be developed by the students):

Through interaction with IHMT researchers, guests from externals to the institution and autonomous research, at the end of the curricular unit of Genetics of Drug Resistance in Parasitology, students should:

1. Know notions of antiparasitic resistance genetics, phenotypic adaptation to the environment and the challenges in defining resistance;
2. Recognize different areas of interest in the study of the genetics of antiparasitic resistance and its relevance;
3. Define the main resistance mechanisms and their molecular basis in the context of medical parasitology;
4. Be aware of notions of pharmacogenetics and drug resistance in parasitology;
5. Understand, analyze and evaluate the applicability of some methodologies and tools for the identification of genetic markers of resistance and their applicability in the monitoring, epidemiology and control of parasites with an impact on human health.

Syllabus:

- I. Introduction to the concept of drug resistance and parasitic infection. Notion of resistance genomics. Notion of phenotypic adaptation to the environment, resistance to antiparasitics, transmembrane transport of xenobiotics/drugs in eukaryotes.
- II. Enzymes of the oxidative stress response system and efflux pumps in response to drugs, using the malaria parasite *Plasmodium falciparum* as an example.
- III. Mechanisms of resistance to insecticides. Define the main resistance mechanisms and their molecular basis. Concept of resistance vs tolerance. Types of resistance (physiological, behavioral).
- IV. Monitoring the occurrence of drug resistance: example *Pneumocystis jirovecii*.
- V. Epidemiological importance of resistance to drugs currently used in the treatment of leishmaniasis.
- VI. Resistance to anthelmintics. Difficulties in defining resistance. Genomics of resistance in nematodes, trematodes and cestodes. Individual and community consequences. *Biomphalaria* spp and *Schistosoma mansoni*: Resistant or susceptible. Host specificity and infection.
- VII. Basic concepts of tools - metabolomics, proteomics and transcriptomics.
- VIII. Post-genomic tools for studying drug resistance: Experimental Design; Bioinformatics; Examples of applications.
- IX. Pharmacogenetics and drug-drug interactions. The notions of pharmacogenetics, pharmacogenomics will be addressed. Drug metabolism phenotypes their causes and impacts on drug resistance selection. Some examples in parasitology.

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Teaching methodologies (including assessment):

Teaching of this CU is based on the lecturing method, translated into theoretical classes; In the demonstrative method, in a practical laboratory class; In the active and interrogative methods applied in theoretical-practical classes and in a seminar class. There will also be tutorial sessions to support the autonomous study and preparation of the seminars.

Student evaluation is based on the students' performance in a written multiple-choice test with 30 questions (0.57 values each) and another three short answer questions (one 0.9 points and the other two 1 value each). The approval in the UC implies: i) the accomplishment of a minimum attendance of 75% of the classes and the achievement of a minimum final classification of 10 values (maximum 20 values) in a written test.

References for consultation / mandatory existence:

- O'Neill PM, Barton VE, Ward SA. The molecular mechanism of action of artemisinin--the debate continues. *Molecules*. 2010 Mar 12;15(3):1705-21.
- Witkowski B, Berry A, Benoit-Vical F. Resistance to antimalarial compounds: methods and applications. *Drug Resist Updat*. 2009 Feb-Apr;12(1-2):42-50. doi:
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- Sá JM, Chong JL, Wellems TE. Malaria drug resistance: new observations and developments. *Essays Biochem*. 2011;51:137-60.
- Witkowski B, Lelièvre J, Barragán MJ, Laurent V, Su XZ, Berry A, Benoit-Vical F. Increased tolerance to artemisinin in *Plasmodium falciparum* is mediated by a quiescence mechanism. *Antimicrob Agents Chemother*. 2010 May;54(5):1872-7.
- Hemingway J, Hawkes NJ, McCarroll L, Ranson H. 2004. The molecular basis of insecticide resistance in mosquitoes. *Insect Biochem Mol Biol* 34: 653-665
- Soderlund DM, Knipple DC. 2003. The molecular biology of knockdown resistance to pyrethroid insecticides. *Insect Biochem Mol Biol* 33: 563-577.
- Brogdon WG, McAllister JC. 1998. Insecticide resistance and vector control. *Emerg Infect Dis* 4: 605-613.
- Exploiting Knowledge on *Leishmania* Drug Resistance to Support the Quest for New Drugs Aya Hefnawy, Maya Berg, Jean-Claude Dujardin, Géraldine De Muylder. *Trends in Parasitology*, **2017**. 33: 162-174.
- *Leishmania* antimony resistance: what we know what we can learn from the field. Aït-Oudhia K, Gazanion E, Vergnes B, Oury B, Sereno D. *Parasitol Res*. **2011**. 109:1225-32

GENETICS OF DRUG RESISTANCE IN PARASITOLOGY

References for consultation / mandatory existence: (continuation)

- Whole genome sequencing of multiple *Leishmania donovani* clinical isolates provides insights into population structure and mechanisms of drug resistance. Downing T, Imamura H, Decuypere S, Clark TG, Coombs GH, Cotton JA, Hilley JD, de Doncker S, Maes I, Mottram JC, Quail MA, Rijal S, Sanders M, Schönián G, Stark O, Sundar S, Vanaerschot M, Hertz-Fowler C, Dujardin JC, Berriman M. *Genome Res.* **2011**. 21:2143-56.
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- Vudriko et al. 2016. Emergence of multi-acaricide resistant *Rhipicephalus* ticks and its implication on chemical tick control in Uganda. *Parasites & Vectors.* 9:4.
- Lopez-Arias et al. 2014. Reduced Efficacy of Commercial Acaricides Against Populations of Resistant Cattle Tick *Rhipicephalus microplus* from Two Municipalities of Antioquia, Colombia. *Environmental Health Insights.* 8:71.
- Abdel-Hamid, A.H.Z, Rawi, S.M. & Arafa, A. F, 2006. Identification of genetic marker associated with the resistance to *Schistosoma mansoni* infection using random polymorphic DNA analysis. *Mem. Inst. Oswaldo Cruz*, **101** (8): 863-868.
- Lockyer, A.E, Jones, C.S, Noble, L.R. & Rollinson, D, 2004. Trematodes and snails: an intimate association. *Canadian J. Zool*, **82** (2): 251
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- Committee for Medicinal Products for Veterinary Use (CVMP) (2016) Reflection paper on anthelmintic resistance. Draft 2. http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2016/04/WC500205608.pdf

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References for consultation / mandatory existence: (continuation)

- Kopp SR, Kotze AC, McCarthy JS, Traub RJ and GT Coleman (2008): Pyrantel in small animal 469 medicine: 30 years on. *Vet Journal*, 178: 177-184.
- Madeira de Carvalho, L.M. (2008) Importância da resistência aos anti-helmínticos a propósito da “Roundtable Nematode Resistance, Atenas, 11-13 de Maio de 2007”. *Acta Parasitológica Portuguesa*, 15 (1/2): 79-91.
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